Stability analysis of quality traits in rice hybrids

PV Padmavathi*, PV Satyanarayana, Lal Ahamed M and N Chamundeswari

Agricultural College, Bapatla - 522101, Andhra Pradesh *E-mail: padmaphd05@gmail.com

ABSTRACT

Fifty two hybrid combinations were evaluated for yield and yield contributing characters over four different agro-climatic zones in Andhra Pradesh, India during dry season 2010-11. Eighteen promising hybrids from all locations viz., Maruteru, Warangal, Jagtial and Ragolu which recorded significant higher yield than check were subjected to analysis for eleven quality characters. The analysis of variance of Eberhart and Russell model indicated the genotypes and environments were significant for all the quality characters except for milling per cent for genotypes indicating the diversity among the genotypes and environments studied. The GE interaction was significant only for head rice recovery, water uptake and kernel elongation ratio and non-significant for remaining characters. The high yielding hybrid APMS 9A x MTU II-143-26-2 was stable for head rice recovery and kernel elongation ratio while APMS 10A x MTU 1071 was stable for kernel elongation ratio, alkali spreading value and amylose content.

Key words: rice hybrid, Eberhart and Russell model, GxE interaction, stability parameters

With the enhanced income levels and changing food habits, breeding rice varieties with preferred grain quality features has become the second most important objective after yield. To meet the market and consumer requirements, milling and cooking quality characteristics are also to be improved. Quality characters are important in varietal development and subsequent adoption at the farm level. Grain quality attributes vary among varieties and production environments. So understanding gene expression in different environments is necessary for improving rice cooking quality traits. Dalvi et al. (2007), Panwar et al. (2008) and Waghmode and Mehta (2011) reported the existence of GE interaction for quality traits of rice. The present study was undertaken to investigate the influence of genotype x environment interaction on grain quality attributes in rice hybrids.

MATERIAL AND METHODS

The experimental material comprised of 52 rice hybrids combination along with three checks [(*viz.*, MTUHR 2089 (hybrid check), MTU 1075 and MTU 1010 (varietal checks)] were evaluated at four different

□ 208 □

agro-climatic zones viz., Regional Agricultural Research Station, Maruteru, Warangal, Jagtial and Agricultural Research Station Ragolu, Andhra Pradesh during dry season 2010-11. Twenty eight days old seedlings were planted at the rate of one seedling hill⁻¹ with a spacing of 20 x15 cm in a randomized block design with two replications at all the locations. Eighteen promising hybrids from all locations which recorded significant higher yield than high yielding check MTU 1075 were subjected to analysis for eleven quality characters (physico- chemical) viz., hulling per cent (H%), milling per cent (M%), head rice recovery (HRR%), L/B ratio, water uptake (WU), volume expansion ratio (VER), kernel elongation ratio (KER), gel consistency (GC), alkali spreading value (ASV), amylose content (AC) and protein content (PC). Physico-chemical characters were recorded by standard evaluation methods (DRR 2006). The biochemical quality parameters viz., gel consistency, alkali spreading value, amylose content and protein content were estimated by using standard evaluation methods of Cagampang et al. (1973), Little et al. (1958), Juliano (1971) and Piper (1966), respectively. Quality analysis data of hybrids and checks from four locations were subjected to pooled stability

analysis as per Eberhart and Russell (1966). The genotype with high mean, unit regression coefficient and non-significant deviation from regression was considered to be stable over environments. If b_i was equal to unity, a genotype was considered to have average stability (same performance in all the environments). If b_i was more than unity, it was suggested to have less than average stability (good performance in favourable environments) and if b_i was less than unity, it was reported to have more than average stability (good performance in poor environments).

RESULTS AND DISCUSSION

Analysis of variance of stability revealed that the genotypes and environments were significant for all the quality characters except for milling percentage indicating the diversity among the genotypes and environments studied (Table 1). The GE interaction was significant only for head rice recovery, water uptake and kernal elongation ratio and non significant for the remaining characters. Significant variation due to environment (linear) was observed for all the eleven quality characters revealing the linear contribution of environmental effects and additive environment variance on these characters. Similar results were reported by Dalvi et al. (2007) for L/B ratio, Panwar et al. (2008) for hulling per cent, milling per cent and head rice recovery per cent, and Waghmode and Mehta (2011) for milling per cent, head rice recovery per cent, L/B ratio, protein and amylose content. The mean sum of squares for pooled deviation was significant for all the characters indicating the non-linear response and unpredictable nature of the genotypes by significantly differing for stability. Significant non-linear responses were observed by Dalvi et al. (2007) for L/B ratio and protein content, Panwar et al. (2008) for hulling per cent, milling per cent and head rice recovery per cent and Waghmode and Mehta (2011) for milling per cent.

For hulling per cent none of the hybrids recorded regression value around unity. Only one hybrid APMS 9A x MTU II -187-6-1-1 exhibited high mean with b_i value close to unity (0.93) and non-significant s²d_i for milling per cent exhibiting average stability. Head rice recovery is most important among milling indices, as the economic value of the rice is determined by percentage of unbroken rice kernel. Two hybrids *viz.*,

| l |
|---|

 Table 1. Analysis of variance for stability performance of quality characters in rice.

| Source | d.f. | d.f. H% | M% | HRR% | L/B | WU | VER | KER | GC | ASV | AC | PC |
|---|------------|----------------------------|-------------|---------------------------|-------------|---|-------------|--------------|--------------|-------------|--------------|-----------|
| Genotypes | 20 | 4.88* | 5.87 | 37.30** | 0.07** | 7184.47 ** | 0.49** | 0.04^{**} | 269.61 ** | 2.95** | 3.18** | 0.23 ** |
| Environments | с | 18.04^{**} | 22.75 ** | 949.92** | 0.02* | 659.61** | 0.57* | 0.06^{**} | 191.40 ** | 0.45** | 11.35** | 3.93 ** |
| Env.+ (Geno. x Env.) | 63 | 3.52 | 4.66 | 61.12** | 0.01 | 203.64 | 0.18 | 0.01^{*} | 38.66 | 0.08 | 1.29* | 0.28 ** |
| Genotypes × environment | 60 | 2.79 | 3.76 | 16.68^{*} | 0.01 | 180.85 | 0.16 | 0.01 | 31.02 | 0.06 | 0.79 | 0.10 |
| Environment (linear) | 1 | 54.13** | 68.25 ** | 2849.75** | 0.05** | 1978.82** | 1.72^{**} | 0.17^{**} | 574.19 ** | 1.36^{**} | 34.05** | 11.79 ** |
| Genotypes ×environment (linear) 20 | r) 20 | 3.05 | 1.86 | 32.33** | 0.00 | 259.53* | 0.18 | 0.01^{*} | 21.77 | 0.06 | 0.80 | 0.09 |
| Pooled deviation | 42 | 2.54** | 4.48 ** | 8.44** | 0.01^{**} | 134.76** | 0.14^{**} | 0.01^{**} | 33.95 ** | 0.06** | 0.74^{**} | 0.09 ** |
| Pooled error | 80 | 1.25 | 1.04 | 1.26 | 0.00 | 21.83 | 0.04 | 0.00 | 6.92 | 0.01 | 0.30 | 0.01 |
| H%= Hulling per cent, M%= Milling per cent, HRR% =Head rice recovery%, L/B = Length/Breadth, WU= Water uptake, VER= Volume expansion ratio, KER= Kernel elonosation ratio GC= Gel consistency ASV= Alkali survadino value AC= Amylose content PC= Protein content | dilling p(| er cent, HRR SV= Alkali | t% =Head r. | ice recovery ⁰ | %, L/B =] | RR% =Head rice recovery%, L/B = Length/Breadth, WU= Wa hi surreading value AC= Amvlose content PC= Protein content | th, WU= W | Vater uptake | , VER= Volun | ne expansio | on ratio, KE | R= Kernel |

Table 2. Stability parameters for quality characters as per Eberhart and Russell model in promising hybrids with checks in rice

| Hybrids | Code | | Н% | | | М% | | | HRR | % | | L/B ra | atio | | WU | | | VER | |
|---------------------------------|---------------|-------|----------------|----------|----------------|----------------|-------------------------------|----------------|----------------|----------|----------------|----------------|----------|----------------|----------------|-------------------------------|----------------|----------------|----------|
| | | | b _i | S^2d_i | \overline{X} | b _i | S ² d _i | \overline{X} | b _i | S^2d_i | \overline{X} | b _i | S^2d_i | \overline{X} | b _i | S ² d _i | \overline{X} | b _i | S^2d_i |
| APMS 6A xMTU II-110-9-1-1-1-1 | A_1R_3 | 77.13 | 1.14 | 1.22 | 67.25 | 1.34 | -0.45 | 56.75 | 0.60 | 1.18 | 2.95 | 2.93 | 0.00 | 106.88 | -0.14 | -14.12 | 3.90 | 0.30 | -0.02 |
| APMS 6A xMTU II -187-6-1-1 | A_1R_5 | 77.88 | 1.35 | -0.95 | 67.63 | 1.35 | -0.28 | 51.88 | 1.48 | 2.91 | 3.17 | 3.47 | 0.01 | 130.00 | 0.67 | 7.11 | 4.47 | 0.24 | -0.03 |
| APMS 6A xMTU II-143-26-2 | A_1R_7 | 78.13 | 1.96 | -0.82 | 67.88 | 1.24 | 2.93 | 53.75 | 1.72 | 0.03 | 3.11 | 0.76 | 0.00 | 110.00 | 0.87 | 17.56 | 4.30 | 0.34 | -0.01 |
| APMS 6A xMTU II-283-7-1-1 | $A_{1}R_{12}$ | 75.75 | 0.44 | -0.33 | 66.88 | 0.39 | 2.69 | 53.38 | 1.38 | 0.43 | 3.06 | 0.87 | 0.02 | 101.88 | -0.26 | -10.22 | 4.25 | 3.64 | 0.48 |
| APMS 9A xMTU 1071 | $A_2 R_1$ | 75.63 | 2.48 | 2.47 | 64.50 | 1.32 | 17.76 | 56.38 | 0.88 | 4.55 | 3.13 | 1.84 | 0.00 | 189.38 | 7.07 | 276.29 | 4.39 | -0.50 | -0.01 |
| APMS 9A xMTU II-110-9-1-1-1-1 | $A_2 R_3$ | 76.50 | 3.69 | 0.45 | 66.88 | 2.46 | 6.86 | 58.13 | 0.68 | 4.74 | 3.08 | 2.67 | 0.00 | 110.00 | 0.36 | 47.09 | 4.09 | 0.50 | 0.13 |
| APMS 9A xMTU II-110-11-1-1-6 | A_2R_4 | 76.88 | 1.35 | 9.55 | 67.75 | 0.65 | 7.54 | 57.25 | 0.64 | -1.19 | 3.03 | -0.42 | 0.00 | 150.63 | 1.80 | 27.17 | 4.26 | 0.77 | 0.23 |
| APMS 9A xMTU II -187-6-1-1 | A_2R_5 | 77.25 | 1.89 | 1.32 | 68.00 | 0.93 | -0.80 | 56.38 | 0.62 | 2.18 | 3.12 | -0.32 | 0.00 | 108.75 | -0.32 | -17.35 | 4.01 | 1.97 | 0.11 |
| APMS 9A xMTU II-143-26-2 | $A_{2}R_{7}$ | 77.63 | 1.31 | -0.59 | 67.75 | 1.19 | -0.83 | 55.75 | 1.03 | 3.65 | 2.90 | 0.83 | 0.01 | 161.38 | 1.94 | 229.50 | 4.65 | 3.27 | 0.15 |
| APMS 9A x MTU II-283-7-1-1 | $A_2 R_{12}$ | 77.13 | -0.04 | 5.88 | 68.63 | -1.15 | 19.57 | 55.13 | 1.05 | 7.86 | 3.17 | -0.12 | 0.01 | 109.38 | 1.17 | 116.13 | 4.05 | 2.75 | 0.16 |
| APMS 10A x MTU 1071 | A_3R_1 | 78.75 | 1.87 | -0.34 | 69.13 | 1.41 | -0.52 | 54.63 | 0.77 | 5.71 | 3.04 | 1.43 | 0.00 | 173.75 | 0.68 | 22.15 | 4.69 | -0.23 | 0.11 |
| APMS 10A x MTU II -187-6-1-1 | A_3R_5 | 78.63 | 0.66 | -0.92 | 68.50 | 0.38 | -1.13 | 53.13 | 1.16 | 7.68 | 3.08 | 1.74 | 0.00 | 163.13 | 1.68 | 41.55 | 4.55 | 0.47 | -0.03 |
| APMS 10A xMTU II-190-1-1-1-1 | A_3R_6 | 77.63 | 0.50 | 0.06 | 68.25 | 0.37 | 0.26 | 49.00 | 1.23 | 44.52 | 2.76 | 0.18 | 0.00 | 130.00 | -0.11 | 302.61 | 5.00 | 2.60 | 0.37 |
| APMS 10A x MTU II-143-26-2 | $A_{3}R_{7}$ | 77.88 | -0.26 | -0.70 | 67.50 | 0.07 | -0.65 | 47.50 | 1.05 | 27.56 | 2.92 | 0.26 | 0.00 | 228.13 | 1.30 | 682.06 | 5.30 | 4.24 | 0.22 |
| APMS 10A x MTU II-290-42-1 | $A_{3}R_{10}$ | 77.25 | -0.39 | 0.72 | 65.13 | 1.50 | 2.31 | 48.75 | 1.16 | 18.41 | 3.09 | 0.36 | 0.00 | 178.13 | 2.24 | 163.01 | 4.32 | 1.75 | 0.21 |
| APMS 10A x MTU II-283-7-1-1 | $A_{3}R_{12}$ | 78.25 | -0.09 | 0.16 | 67.00 | 1.09 | 4.17 | 52.13 | 1.79 | 3.96 | 3.01 | 0.26 | 0.00 | 168.75 | 0.41 | 254.77 | 3.97 | -0.34 | 0.05 |
| IR 58025A xMTU II-110-9-1-1-1-1 | A_4R_3 | 75.88 | 2.00 | -0.53 | 66.38 | 2.20 | 1.58 | 51.13 | 1.43 | -0.81 | 3.33 | 0.87 | 0.00 | 159.38 | 1.17 | 116.13 | 4.00 | -0.41 | 0.02 |
| IR 58025A xMTU II-190-1-1-1-1 | $A_4 R_6$ | 75.38 | -0.40 | 9.68 | 66.88 | 0.75 | 3.52 | 49.50 | 1.60 | 3.47 | 3.24 | -0.26 | 0.00 | 216.25 | -1.22 | 67.75 | 4.31 | -0.26 | 0.05 |
| MTUHR 2089 | | 74.63 | 1.33 | -0.90 | 64.75 | 1.02 | 1.53 | 50.38 | -0.18 | 2.18 | 3.11 | -0.37 | 0.03 | 156.25 | -0.14 | 11.63 | 4.50 | 0.00 | -0.04 |
| MTU 1075 | | 76.75 | -0.24 | 0.85 | 67.00 | 1.03 | -1.11 | 53.88 | 0.41 | 2.57 | 3.14 | 2.03 | 0.00 | 151.88 | 0.05 | 17.89 | 4.34 | -0.68 | -0.02 |
| MTU 1010 | | 77.38 | 0.45 | 1.62 | 66.75 | 1.44 | 5.10 | 54.38 | 0.50 | 8.17 | 2.90 | 1.98 | 0.00 | 253.75 | 1.78 | 12.54 | 4.06 | 0.57 | -0.03 |
| General mean | | 77.06 | | | 67.16 | | | 53.29 | | | 3.06 | | | 155.13 | | | 4.35 | | |

Table 2 contd.

Table 2 contd.

| Hybrids | | KER | | | œ | | | ASV | | | AC | | | PC | |
|---------------------------------|----------------|----------------|----------|----------------|----------------|-------------------------------|----------------|----------------|----------|----------------|----------------|-------------------------------|----------------|----------------|----------|
| | \overline{X} | b _i | S^2d_i | \overline{X} | b _i | S ² d _i | \overline{X} | b _i | S^2d_i | \overline{X} | b _i | S ² d ₁ | \overline{X} | b _i | S^2d_i |
| APMS 6A x MTU II-110-9-1-1-1-1 | 1.67 | 2.22 | 0.02 | 75.25 | -0.35 | 62.33 | 2.20 | 0.19 | 0.00 | 24.75 | -0.20 | 1.97 | 8.25 | 2.07 | 0.14 |
| APMS 6A x MTU II -187-6-1-1 | 1.68 | 0.61 | 0.00 | 74.75 | 1.17 | -4.60 | 2.38 | 0.08 | 0.00 | 23.47 | 0.76 | 0.72 | 8.54 | 0.83 | -0.01 |
| APMS 6A x MTU II-143-26-2 | 1.65 | 1.73 | 0.00 | 70.09 | 1.33 | -2.78 | 2.33 | 0.45 | 0.04 | 24.08 | 1.80 | 0.04 | 8.48 | 1.10 | 0.12 |
| APMS 6A x MTU II-283-7-1-1 | 1.86 | 0.64 | 0.00 | 82.49 | -0.48 | 16.42 | 2.18 | 0.48 | 0.00 | 25.28 | 1.95 | 0.01 | 8.20 | 0.77 | 0.01 |
| APMS 9A x MTU 1071 | 1.75 | -0.72 | 0.01 | 65.00 | 0.58 | 0.81 | 2.87 | 2.63 | 0.44 | 25.69 | -0.15 | -0.21 | 8.07 | 0.59 | 0.15 |
| APMS 9A x MTU II-110-9-1-1-1-1 | 1.71 | -1.32 | 0.00 | 66.38 | 0.88 | -2.27 | 2.16 | 0.94 | 0.01 | 23.67 | 0.92 | 0.37 | 8.27 | 0.91 | 0.17 |
| APMS 9A x MTU II-110-11-1-1-6 | 1.75 | 0.04 | 0.00 | 82.13 | 1.21 | 5.87 | 2.20 | 1.17 | 0.01 | 23.91 | 1.01 | -0.26 | 8.40 | 0.43 | 0.08 |
| APMS 9A x MTU II -187-6-1-1 | 1.75 | 2.10 | 0.00 | 75.50 | -0.30 | 88.59 | 2.08 | 0.48 | 0.00 | 23.37 | 1.40 | 0.03 | 7.91 | 0.67 | 0.03 |
| APMS 9A x MTU II-143-26-2 | 1.81 | 1.00 | 0.01 | 66.97 | 1.54 | -7.14 | 2.86 | 0.57 | 0.05 | 24.65 | 0.58 | -0.09 | 8.03 | 0.90 | 0.00 |
| APMS 9A x MTU II-283-7-1-1 | 1.82 | 1.69 | 0.00 | 69.38 | 1.89 | 14.84 | 2.16 | 0.71 | 0.00 | 25.10 | 1.85 | 0.54 | 8.09 | 1.16 | 0.03 |
| APMS 10A x MTU 1071 | 1.90 | 1.02 | 0.00 | 73.63 | 1.34 | -5.42 | 4.18 | 0.94 | 0.05 | 24.14 | 0.85 | -0.12 | 8.57 | 1.54 | 0.15 |
| APMS 10A x MTU II -187-6-1-1 | 1.82 | 1.76 | 0.01 | 72.63 | -0.45 | 73.96 | 3.09 | 3.77 | 0.06 | 23.77 | 1.24 | 0.51 | 8.04 | 1.08 | 0.09 |
| APMS 10A x MTU II-190-1-1-1-1-1 | 1.94 | 1.65 | 0.00 | 72.75 | 0.95 | -5.37 | 2.54 | 0.31 | 0.28 | 24.15 | 1.90 | 2.76 | 8.11 | 1.17 | 0.07 |
| APMS 10A x MTU II-143-26-2 | 1.87 | -0.28 | 0.01 | 68.25 | 1.56 | 64.80 | 4.29 | 0.82 | 0.09 | 24.37 | 0.76 | 0.14 | 7.99 | 0.83 | 0.02 |
| APMS 10A x MTU II-290-42-1 | 1.76 | -0.54 | 0.00 | 44.25 | 1.50 | -4.72 | 4.22 | 0.36 | 0.00 | 24.16 | 1.72 | 0.45 | 7.86 | 0.95 | 0.11 |
| APMS 10A x MTU II-283-7-1-1 | 1.96 | 1.28 | 0.00 | 82.25 | -0.11 | 29.80 | 3.62 | 0.11 | 0.03 | 24.17 | 0.68 | -0.26 | 8.36 | 0.26 | -0.01 |
| IR 58025A x MTU II-110-9-1-1-1 | 1.64 | 4.57 | 0.00 | 79.00 | 2.28 | 14.00 | 2.28 | 0.67 | 0.02 | 23.26 | 1.54 | 0.59 | 7.98 | 1.54 | 0.07 |
| IR 58025A x MTU II-190-1-1-1-1 | 1.92 | 1.47 | 0.01 | 76.25 | 1.39 | 143.96 | 4.19 | 0.67 | 0.02 | 23.97 | -0.12 | 0.64 | 8.13 | 1.14 | -0.01 |
| MTUHR 2089 | 1.77 | 0.49 | 0.00 | 77.00 | 2.49 | 44.95 | 2.88 | 1.62 | 0.00 | 24.54 | 0.12 | 0.00 | 7.73 | 1.33 | 0.43 |
| MTU 1075 | 1.76 | 0.45 | 0.00 | 70.70 | 0.76 | 26.20 | 2.88 | 1.62 | 0.00 | 23.88 | 0.71 | 1.10 | 8.21 | 0.73 | 0.03 |
| MTU 1010 | 1.73 | 1.17 | 0.00 | 70.38 | 1.82 | 8.59 | 4.61 | 2.42 | 0.05 | 21.28 | 1.68 | 0.55 | 8.58 | 0.99 | 0.01 |
| General mean | 1.79 | | | 72.14 | | | 2.96 | | | 24.08 | | | 8.18 | | |

 \overline{X} = mean, b_i = regression coefficient, S²d_i = deviation from , H%= Hulling per cent, M%= Milling per cent, HRR% =Head rice recovery%, L/B = Length/Breadth, WU= Water uptake, VER= Volume expansion ratio, KER= Kernel elongation ratio, GC= Gel consistency, ASV= Alkali spreading value, AC= Amylose content, PC= Protein content

Quality traits in rice hybrids

APMS 9A x MTU II-143-26-2 ($b_i=1.03$) and APMS 9A x MTU II-283-7-1-1 ($b_i=1.05$) were recorded to have mean higher than grand mean, b_i values around unity and non significant s²d_i values for head rice recovery per cent (Table 2). Dalvi *et al.* (2007) and Waghmode and Mehta (2011) recorded stable hybrids for milling per cent and head rice recovery. Panwar *et al.* (2008) reported ideal genotypes with general adaptability for hulling per cent.

The hybrid IR 58025A x MTU II-110-9-1-1-1-1 ($b_i=0.87$) possessed average stability for L/B ratio with b_i value around unity and zero s²d_i whereas two hybrids, APMS 9A x MTU II-143-26-2 ($b_i=1$) and APMS 10A x MTU 1071 ($b_i=1.02$) showed unit regression coefficient values ($b_i=1$) with above grand mean and non-significant deviation from linearity (s²d_i=0) for kernel elongation ratio, reveals the stability of genotypes across environments. Similar findings of stable hybrids were reported by Dalvi *et al.* (2007) and Waghmode and Mehta (2011) for L:B ratio.

Gel consistency gives softness of rice and soft to medium gel consistency is preferred one by most of the rice consumers (Siddiq, 1992). For this trait average stability exhibited by APMS 10A x MTU II-190-1-1-1-1-1 hybrid with high mean, b, value (b=0.95) around unity and non significant s²d, value while, APMS 10A x MTU 1071 hybrid recorded regression value around unity (0.94) with above general mean and nonsignificant deviation from regression for alkali spreading value and was found to be stable under four environments. Amylose content of rice determines the hardness or stickiness of cooked rice. Average stability for this trait was possessed by APMS 9A x MTU II-110-11-1-1-6 (b_i=1.01) and APMS 9A x MTU II-110-9-1-1-1 (b=0.92) hybrids with low mean than general mean and regression value around unity with predictable performance. One hybrid APMS 9A x MTU II-110-9-1-1-1 ($b_i=0.91$) and check MTU 1010 (b=0.99) registered regression values around unity with above grand mean and predictable performance for protein content and were found to be stable over locations. Waghmode and Mehta (2011) reported stable hybrids for amylose content and protein content whereas Dalvi et al. (2007) found hybrids with average stability for the trait protein content.

Environmental index (I_j) reveals the suitability of an environment. Based on the positive values of environmental index for grain yield. Maruteru (I=5.06) and Warangal (I_i =4.66) locations were found to be most suitable environments compared to Jagtial (I=2.56), while Ragolu (I = -12.28) location was poor in performance. Six hybrids each were adapted to favourable (better condition) and unfavourable (poor condition) environments for hulling per cent. For milling per cent, five hybrids each specifically adapted to better and poor environment conditions, whereas for head rice recovery per cent two and six hybrids where found to be ideal under favourable and unfavourable environments, respectively. For the trait L/B ratio four hybrids were suitable under favourable environments and five hybrids for unfavourable environments (Table 3). Panwar et al. (2008) and Waghmode and Mehta (2011) also reported genotypes to favourable and unfavourable environments for these traits.

Six hybrids were found to be adapted in better condition while three hybrids in poor conditions for water uptake. For volume expansion ratio, three hybrids were suitable to favourable and four hybrids for unfavourable environments. Whereas five hybrids were found to be ideal under favourable environment and two hybrids were suitable to unfavourable environment for kernel elongation ratio. With regards to gel consistency, it was observed that five hybrids were found to be highly responsive under favourable environment whereas four hybrids were found to be ideal for unfavourable environment. One hybrid was ideal in favourable environment for alkali spreading value and four hybrids were adapted in unfavourable environment for this trait. Two hybrids were suitable under better environment for amylose content, while three hybrids recorded ideal performance under unfavourable environment. For protein content, three hybrids were found to be specifically adapted to better environment while four hybrids were suitable to poor environments. Similar findings of genotypes to favourable and unfavourable environments for the traits viz., amylose content and protein content were also reported by Panwar et al. (2008) and Waghmode and Mehta (2011).

The hybrid, APMS 9A x MTU II-143-26-2 with high yield was stable for head rice recovery and kernel elongation ratio while APMS 10A x MTU 1071 had stable performance for kernel elongation ratio, alkali spreading value and amylose content. From the present

Oryza Vol. 50 No.3, 2013 (208-213)

| Character | Stable performance | Favourable environment | Poor environment |
|---|--|---|--|
| Hulling per cent | — | $A_1R_3, A_2R_7, A_1R_5, A_3R_1, A_2R_5 \text{ and} A_1R_7$ | , A_3R_5 , A_3R_6 , A_2R_{12} , A_3R_{12} , $A_3R_7A_3R_{10}$ and checks MTU 1010 and MTU 1010 |
| Milling per cent | A_2R_5 | A_2R_7 , A_1R_7 , A_1R_3 , A_1R_5 and A_3R_1 | A_2R_4 , A_3R_5 , A_3R_6 , A_3R_7 and A_2R_{12} |
| Head Rice Recovery per cent | $A_2 R_7$ and $A_1 R_{12}$ | A_1R_{12} and A_1R_7 | $A_2R_1, A_3R_1, A_2R_3, A_2R_4, A_1R_3, A_2R_5$ and checks MTU 1075 and MTU 1010 |
| L/B Ratio | A_4R_3 | A_3R_5 , A_2R_1 , A_2R_3 , A_1R_5 and check MTU 1075 | $A_1R_7, A_3R_{10}, A_2R_5, A_2R_{12} \mbox{ and } A_4R_6 \mbox{ and } check \mbox{ MTUHR } 2089$ |
| Water uptake | _ | $A_4 R_3,\ A_3 R_7,\ A_3 R_5, A_2 R_7,\ A_3 R_{10},\ A_2 R_1$ and check MTU 1010 | A_3R_1 , A_3R_{12} , A_4R_6 and check MTUHR 2089 |
| Volume expansion ratio | | A_3R_6 , A_2R_7 and A_3R_7 | $A_{3}R_{5}$, $A_{1}R_{5}$, $A_{3}R_{1}$, $A_{2}R_{1}$ and MTUHR 2089 |
| Kernal elongation ratio | A_2R_7 and A_3R_1 | $A_{3}R_{12}, A_{4}R_{6}, A_{3}R_{6}, A_{2}R_{12}$ and $A_{3}R_{5}$ | A_1R_{12} and A_3R_7 |
| Gel consistency | A_3R_6 | $A_1R_5, A_2R_4, A_3R_1, A_4R_6, A_4R_3$ and check MTUHR 2089 | A_2R_5 , A_1R_3 , A_3R_5 and A_1R_{12} |
| Alkali spreading value | A_3R_1 | $A_{3}R_{5}$ and check MTU1010 | $A_{3}R_{7}$, $A_{4}R_{6}$, $A_{3}R_{10}$ and $A_{3}R_{12}$ |
| Amylose content | A_2R_4 and A_2R_3 | A_3R_5 , A_2R_5 , A_4R_3 hybrids and check MTU 1010 | A_2R_4 , A_1R_5 , A_4R_6 and check MTU 1075 |
| Protein content | A_2R_3 and MTU 1010 | $A_1R_7 A_3R_1$ and A_1R_3 | $A_1R_5,\ A_1R_{12},\ A_2R_4$, A_3R_{12} and check MTU 1075 |
| $\overline{A_1R_3}$ -APMS 6A xMTU II-110- A_1R_{12}-APMS 6AxMTU II-283- A_2R_4-APMS 9AxMTU II-110-1 A_2R_{12}-APMS 9AxMTU II-283- A_3R_6-APMS 10A xMTU II-283 A_3R_12-APMS 10A xMTU II-283 | $\begin{array}{cccc} 7\text{-}1\text{-}1 & A_2 R_1 \\ 11\text{-}1\text{-}1\text{-}1\text{-}6 & A_2 R_5 \\ 7\text{-}1\text{-}1 & A_3 R_1 \\ -1\text{-}1\text{-}1\text{-}1\text{-}1 & A_3 R_7 \end{array}$ | -APMS 9AxMTU 1071 -APMS 9A xMTU II -187-6-1-1 APMS 10AxMTU 1071 APMS 10AxMTU II-143-26-2 | $\begin{array}{l} A_1 R_7 \text{ -} APMS \ 6AxMTU \ II-143-26-2 \\ A_2 \ R_3 \text{ -} APMS \ 9AxMTU \ II-110-9-1-1-1-1 \\ A_2 R_7 \text{ -} APMS \ 9A \ xMTU \ II-143-26-2 \\ A_3 R_5 \text{ -} APMS \ 10A \ x \ MTU \ II \ -187-6-1-1 \\ A_3 R_{10} \text{ -} APMS \ 10A \ x \ MTU \ II \ -290-42-1 \\ A_4 R_6 \text{ -} IR \ 58025AxMTU \ II-190-1-1-1-1-1 \end{array}$ |

Table 3. Stable hybrids for quality characters as per regression model of Eberhart and Russell in rice

study it can be concluded that the hybrids *viz.*, APMS 9A x MTU II-143-26-2 and APMS 10A x MTU 1071 were stable for important cooking quality characters with high grain yield plant⁻¹ which may be exploited for commercial cultivation in rice growing areas of Andhra Pradesh.

REFERENCES

- Cagampangde GB, Pere ZCM and Juliano BO 1973. A gel consistency test for eating quality of rice. *J. Sci. Fd Agric.*, 24: 1589-1594.
- Dalvi VV, Patel DV, Vashi PS and Khirsagar RM 2007. Genotype x environment interaction for yield and its components in rice hybrids. J. Maharastra Agric. Univ., 32 (1) 25-28.
- Directorate of Rice Research (DRR, Hyderabad). 2006. Laboratory Manual on Rice Grain Quality Procedures.
- Eberhart SA and Russell WA 1966. Stability parameters for comparing varieties. Crop Sci., 6: 36-40.

- Juliano BO 1971. A simplified assay for milled rice amylose. Cereal Sci. Today. 16 (10): 334-339.
- Kempthorne O 1957. An Introduction to Genetic Statistics. John Wiley and Sons Inc, New York pp: 458-471
- Little RR, Hilder GB and Dawson EH 1958. Differential effect of dilute alkali on 25 varieties of milled white rice. Cereal Chem., 35:111-126.
- Panwar LL, Joshi VN and Mashiat Ali 2008. Genotype x environment interaction in scented rice. Oryza, 45 (1): 103-109.
- Piper CS 1966. Soil and Plant Analysis. Inter Science Publishers, New York. 47-49.
- Siddiq E A 1992. Rice production strategy for the 21st century. Third Dr. K. Ramiah Memorial lecture held on 26th August 1992, at Orissa University of Agriculture and Technology, Bhuaneshwar.
- Waghmode BD and Mehta HD 2011. Genotype x environment interaction and stability analysis in hybrid rice. Crop Improv. 38 (1): 6-12.